

Comments on Biofilm Microbiology as distinct from Planktonic Microbiology

With implications for human Bacterial Endocarditis:

By Alan B. MacDonald MD, FCAP, FASCP

Dr. Bill Costerton practices outside of the perimeter of Planktonic Microbiology.

We Hospital pathologists were certified by the American Board of Pathology. Those who opt to Sit for the examination in Clinical Pathology achieve Certification in Microbiology. "Planktonic" Microbiology is now the correct modifier for this certification status because there are no American Board of Pathology Examination questions in Biofilm Science. So graduates of the usual Hospital clinical pathology training programs are uneducated in Biofilm type Microbiology. Biofilm microbiology operates Under a set of teachings which is foreign to Traditional Microbiology training at either the Medical Doctoral level (MD or DO) ; or at the Doctor of Philosophy level.(PhD). Dr. Bill Costerton does have a following. I am a convert. Biofilms explain persistence of viable microbes in chronic infections, and all infections of medical prostheses.

I am very interested in persistence of *Borrelia burgdorferi* infections in the human host. Biofilms are communities with their own "... self generated Extracellular matrix protective shield," and their own electrical nano-communication systems, and their own capricious behavior to break off and embolize to distant sites, or to "shower disseminate Planktonics from the biofilm community into the body fluids.

Costerton says , to paraphrase, that conventional microbiology is academically Great, but shackled by the " Microbiology nutrient agar plate type evaluation ". Academic microbiology with its focus on Planktonics is ill equipped to deal with biofilm medicine. Dr. Stephen Barthold's favorite "VBNC" microbes; {Viable But Non Cultivable(s)} add yet another reason as to why a living borrelia in mammalian biofilm community within living tissue would FAIL to grow in BSKH under conditions of Maximal Planktonic Support.

Biofilm communities, when explanted from human tissues (i.e. Infected hip prostheses for instance), will not grow on agar plates in a hospital laboratory. Costerton points out in his bibliography of 600 peer reviewed manuscripts and book chapters that Biofilm Science does not pretend to be governed by purely planktonic principles.

Internet Link: Dr. Bill Costerton - The "Father" of Biofilms - on YouTube Internet site
:http://www.youtube.com/watch?v=M_DWNFFgHbE

In the Biofilm of *Borrelia burgdorferi* community, as my Hyper Spectral High Resolution Images from the CYTOVIVA research apparatus, demonstrate that specialization of microbes within a biofilm community [Non spiral forms, granular forms, cystic forms, and cell wall deficient forms].

is the rule, and part of that specialization is biochemical shifts in metabolic requirements, and indeed biochemical shifts in the constituents of cell wall structure and microbial shapes among the members of that community. Spiral bacteria (Leptospire =Biofilm formers, Oral Treponemes=biofilm formers) and now at long last *Borrelia* species= Biofilm formers; all of these undergo shape shifting, while maintaining viability.

Granular forms of *borrelia burgdorferi* are perhaps contentious entities for some career *borrelia* investigators to accept as bona fide viable replicating forms of a microbial life form which is “supposed” to be only spiral in profile. Cystic forms, [notwithstanding the omnibus of the works of the Drs. Brorson,] did not find a place in an encyclopedic excellent (and justifiably expensive) monograph on *Borrelia* published in year 2011. Cell wall deficient (spheroplast) forms have been documented by some researchers (funded for study at Johns Hopkins Medical School) but are rejected by many career *borrelia* researchers in active practice and in positions of Editorial power in the USA and European research communities. So morphologic diversity- legitimate, viable, part of a spirochete life cycle - are topics for a future monograph on *borrelia* microbes .I attach some very old work from Dr. Elisabeth Aberer and Dr. Paul Harrison Duray, and from Dr. Edward Delamater, and for historical interest.

Biofilm infections are the “stuff” of infections of the human heart valves (Bacterial endocarditis). In Bacterial Endocarditis, sessile biofilm communities attach themselves to the surface of the human Endothelial lined heart valves, and produce “vegetations” or “bumpy fibrin covered distortions” of the profiles of a healthy smooth surfaced delicate thin heart valve leaflet or cusp. Over time, the Biofilm communities which grow in size, are detectable by Radiologic imaging of the heart valves. The Heart valves are injury or in some cases completely destroyed by the biofilm communities of infecting microbes which have become attached to the valvular surface.

Over time, with persistence of the biofilm infection of the heart valve surface, bits of the Biofilm community break off from the parent biofilm unit and embolize (spread to distant Body sites). In addition, some biofilm infections of heart valves send out Showers “Seeding dissemination” of Planktonic bacteria into the blood stream. New sites of biofilm infection are thus established In the body of the host human with bacterial endocarditis.

In year 2012, the first report of Human Bacterial Endocarditis was reported from France.

Article (full text) reproduced below The Image within the article illustrates the microscopic profile of the Vegetation which was attached to the diseased heart valve. Attempts to visualize spiral (planktonic) *Borrelia burgdorferi* within the vegetation disclosed no spiral shaped microbes using a silver stain.

Lyme endocarditis

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Abstract

Lyme borreliosis is a common tick-borne disease with a wide variety of clinical manifestations. Cardiac involvement has been reported during both the acute phase (atrioventricular block, pericarditis) and the chronic stage (dilated cardiomyopathy), but is rare (<5%). Here we describe the first case of *Borrelia afzelii* Lyme endocarditis, in a 61-year-old man living in an endemic area of France. The diagnosis was confirmed by detection of *B. afzelii* DNA in the mitral valve by specific real-time PCR. He was treated empirically with amoxicillin for 6 weeks and remains well 12 months later.

Keywords: *B. afzelii*, *Borrelia*, borreliosis, endocarditis, lyme

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Lyme borreliosis (or Lyme disease) is the most commonly reported tick-borne disease in the northern hemisphere, notably in Europe and North America. The different species of the *Borrelia burgdorferi* sensu lato group are transmitted by infected ticks of the genus *Ixodes*. Whereas only one bacterial species, *B. burgdorferi* sensu stricto, is currently recognized as pathogenic in North America, several pathogenic species are present in Europe (mainly *B. burgdorferi* sensu

stricto, *B. afzelii* and *B. garinii*), where they cause a wider variety of clinical manifestations [1]. Typically, following initial erythema migrans at the tick bite site, these bacteria can spread from the skin to other tissues and organs, causing more severe manifestations such as arthritis and cutaneous and neurological disorders [2]. Cardiac Lyme borreliosis is rare, representing only 0.3–4% of cases in Europe, and is generally associated with acute-onset atrioventricular (I–III) conduction disorders, arrhythmias and, in some cases, myocarditis or pericarditis [3–8]. Here we describe a documented case of Lyme endocarditis.

A 61-year-old man was admitted in March 2011 to Limoges University Hospital, France, for mitral valve replacement. He was an ex-smoker, had a history of paroxysmal atrial fibrillation, and had mitral insufficiency due to mitral valve prolapse. Initial investigations showed auricular fibrillation and dyspnoea, mitral regurgitation with rope rupture, an ejection fraction of 45%, and a dilated left atrium on cardiac ultrasound. During surgery the macroscopic aspect of the mitral valve suggested endocarditis, with prolapse of the posterior valve and a 5-mm² perforation of the anterior valve. All blood cultures and serologies commonly performed in the case of endocarditis (*Bartonella henselae*, *Bartonella quintana*, *Mycoplasma pneumoniae*, *Legionella pneumophila*, *Chlamydia pneumoniae* and *Coxiella burnetii*) were all negative. He was treated empirically with intravenous amoxicillin and gentamicin for 2 weeks, followed by oral amoxicillin for 4 weeks. Microscopic analysis of the mitral valve showed endocarditis with foamy macrophages suggestive of intracellular microorganisms (Photo 1). Gram, PAS and Gimenez stains were negative. Whartin-Starry stain showed only

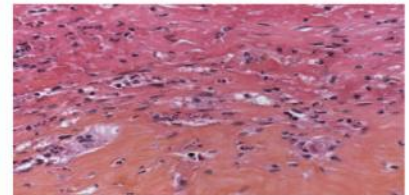


PHOTO 1 Microscopic view of the mitral valve showing surface fibrin deposit, and sparse inflammatory infiltrate consisting of foamy macrophages, neutrophils and eosinophils (hematein eosin saffron $\times 400$).

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scarce curved rods, which had a morphology that was not specific to Spirochaetes. Universal PCR targeting 16S RNA-encoding DNA was applied to a valve fragment and the amplification product was sequenced, identifying the genus *Borrelia*. Two valve fragments were sent to the French Borrelia National Reference Center for confirmation. Both were positive by specific real-time PCR using a Taqman[®] probe targeting a conserved region of the flagellin (*fla*) gene of the *Borrelia burgdorferi* sensu lato (BbsI) complex [9]. Further real-time DNA amplification using hybridization probes targeting species-specific regions of the *fla* gene identified *B. afzelii*. The Anti-*Borrelia* Plus VisE ELISA IgG assay was positive and the Anti-*Borrelia* ELISA IgM assay was negative (both tests from Euroimmun AG, Luebeck, Germany). Western blot (*Borrelia afzelii* + VisE Eco Blot IgG Western Blot; Virotech, Rüsselsheim, Germany) confirmed the presence of several antibodies targeting the VisE, p83, p58, p39, p31 and p21 proteins. Only atrial fibrillation persisted after antibiotic treatment, with no mitral regurgitation. As the patient was well, and given the lack of specific therapeutic guidelines for Lyme endocarditis, antibiotic treatment was not prolonged or changed. The patient did not recall a tick bite, a previous episode of erythema migrans, or secondary manifestations such as meningoradiculitis.

To our knowledge this is the first documented case of *B. afzelii* Lyme endocarditis. In Europe, *B. afzelii* is commonly associated with neurological and late cutaneous manifestations, and less frequently with arthritis. Only one previous publication mentions detection of the weakly pathogenic species *B. bisetii* in a patient's cardiac valve tissue, in the Czech Republic, without mentioning Lyme endocarditis diagnosis [10]. Borreliosis is endemic in our patient's home region (Limousin), with an estimated seroprevalence of 42 cases per 100 000 inhabitants [11]. *B. burgdorferi* seropositivity is very common in endemic areas, and cannot serve as a definitive diagnostic criterion, as underlined by Kaell et al. [12] and Stanek et al. [2]. The exceptional case of Lyme endocarditis described here emphasizes the need to perform universal PCR on heart valve samples in the case of endocarditis of unknown origin, and to bear in mind the possibility of bacterial aetiology in endemic areas.

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Authors' Contributions

NH collected all results; OB wrote the report; SDM, FG and CM carried out microbiological analyses; SS and ML managed the patient; FP carried out microscopic analyses; BJ and MCP reviewed the report.

Conflict of Interest

Written consent to publish was obtained. The authors declare no conflicts of interest.

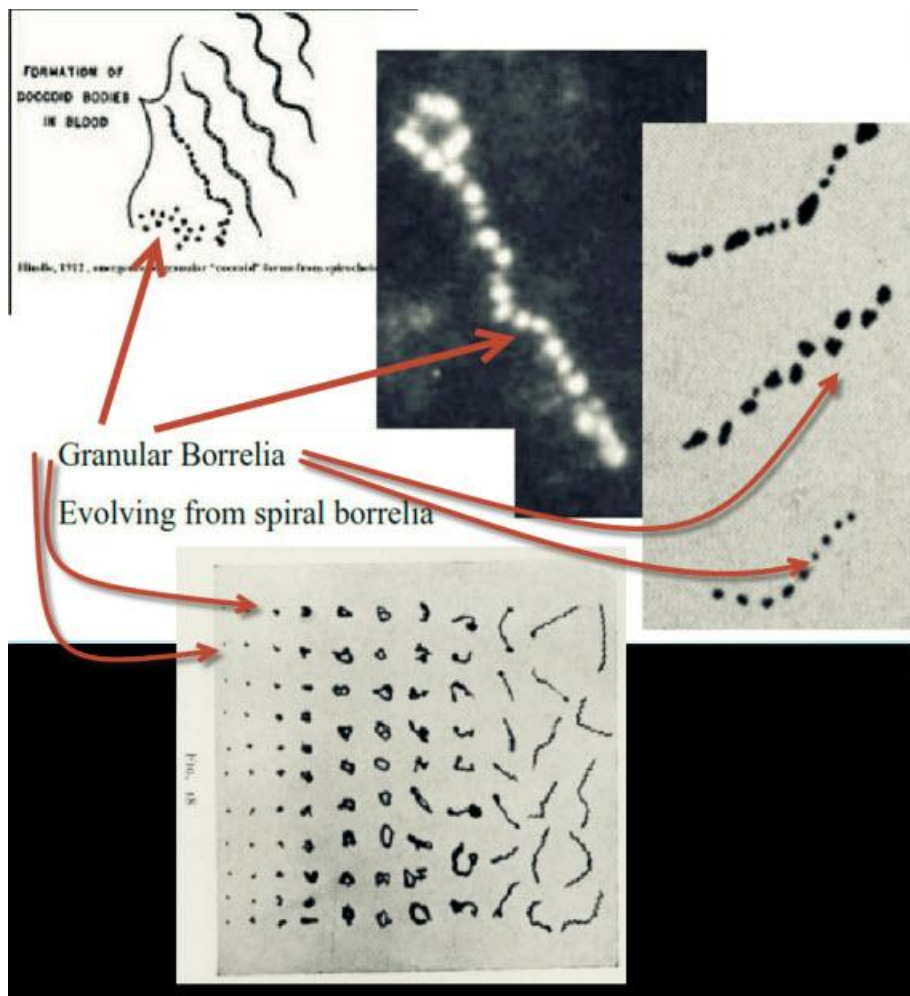
Transparency Declaration

The authors declare no conflicts of interest.

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A pertinent microscopic observation within the silver stained heart valve vegetation using the WarthinStarry silver stain was to observation of “scarce curved rods which had a morphology that was not specific to Spirochetes”. This is indeed a pertinent microscopic observation. The work of Dr. ElisabethAberer And Dr. Paul Harrison Duray (below) illustrates that bona fide spirochetes in controlled Laboratory conditions, often show profiles which are Not Spiral, but indeed may show the Profile of “curved rods”. Lack of awareness of this peer reviewed manuscript from Year 1991 Is an extreme disadvantage to formulating a correct tissue Pathology diagnosis of Borrelia infection in tissue . TheMolecular studies in this case of heart valve tissue which was surgically removed, rigorously confirm thatThe DNA of Borrelia burgdorferi group SI (B. Afzelii) was resident in the diseased and resected heart valve tissues..



**Shape Shifting : Routinely observed by Expert Borrelia Research Pathologists
Elizabeth Aberer MD (Germany) and
Paul Harrison Duray MD (Yale School of Medicine)**

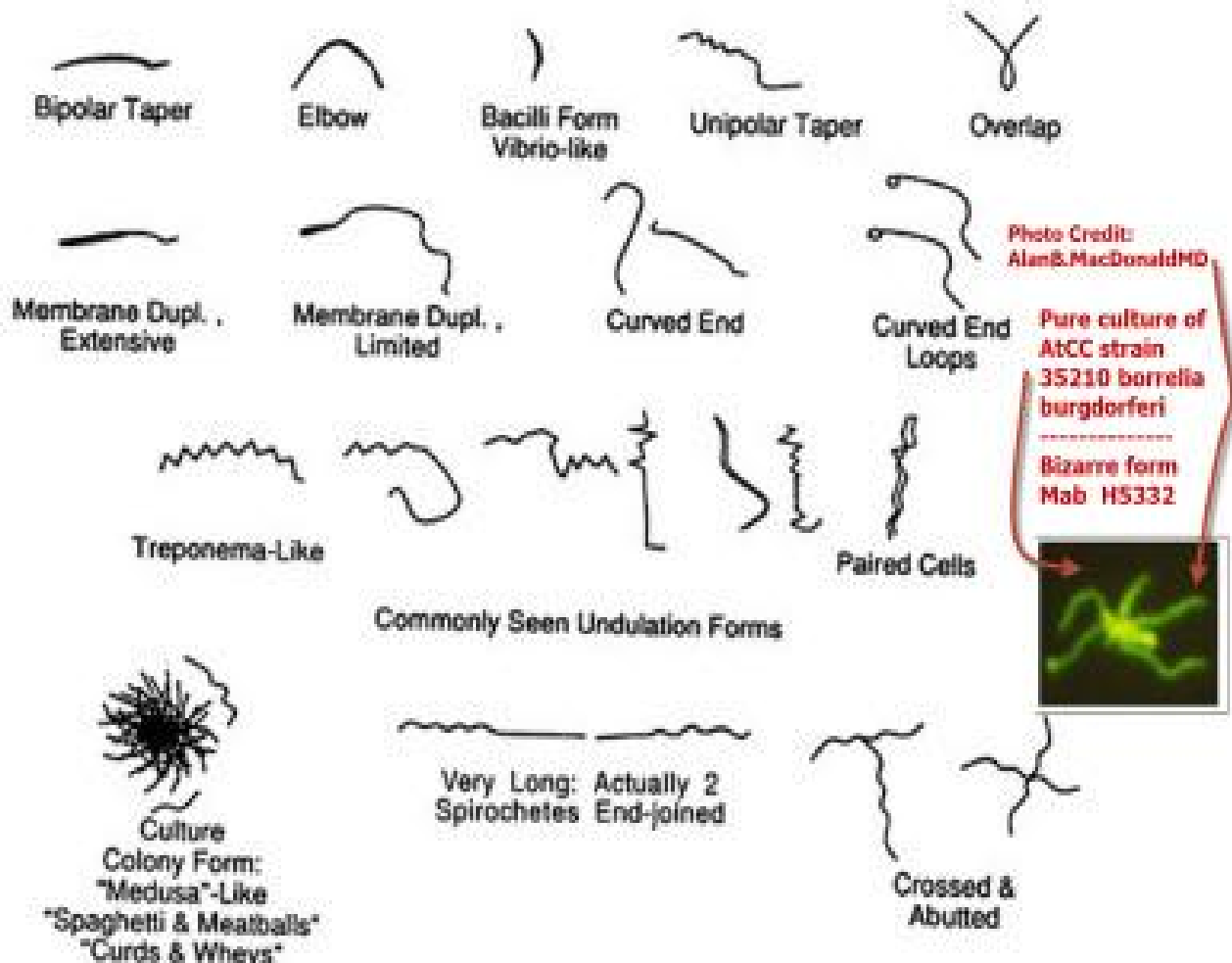


FIG. 1. Common morphologies of *B. burgdorferi* B31.

J Clin Microbiol 1991; 24(2):794-72

Morphology of *Borrelia burgdorferi*: structural patterns of cultured borreliae in relation to staining methods.

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Shape shifting in borrelia burgdorferi::

Non Corkscrew shaped forms:

Straightened forms

Ring forms

Crossed and Abutted Forms

Granular forms

Cystic forms

Cell Wall Deficient forms

Membrane duplicated (ameboid) forms

Shrunk and Collapsed forms (Non-spiral)